

INCORPORATING ESSENTIAL FATTY ACIDS IN SANA WITH FISH OIL ADDED

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Abstract

In this study we have determined the optimal concentrations of fish oil added to raw milk on which the optimal embedding fish oil globule of fat in the product. to this end was made from sheep's milk homogenization temperature of 70 ° c and pressure of 200 bar. The fat globule membrane scission occurs when fat molecule occurs accession fat fish oil to milk fat globules and restoration, which is the number greater use of milk protein composition. we used sheep milk and were analyzed three most important essential fatty acids: linoleic acid, linolenic acid and γ -linolenic acid. Fatty acid analysis was performed by gas chromatography and statistical analysis method of embedding threshold comparison method roc curve (receiver operator characteristic = operating characteristic) and have obtained the following theoretical threshold values potting: linoleic acid, not potting reaches concentrations up to 0.15% fish oil added to milk; 0.650% linolenic acid; 1.319% γ -linolenic acid.

Key words: fish oil essential fatty acids

INTRODUCTION

Consumer interest for products in dairy goats and sheep, especially traditional ones is increasing (Park at Haenlein, 2006). Sheep milk is used in the production has biologically important qualities for man with role in lowering body fat (Versiani and others, 2008). It is known that sheep milk is important not only for liver metabolism but also to cover the manganese is used as food for recovery after illness. It is known that people who practice grazing in several regions of the world have a higher life expectancy (Hanreich et al, 2008).

Because fermentation occurring were identified in products are made up of lactic acid bacteria *Enterococcus faecalis* (Muguerza, B, 2006), *Lactococcus lactis* subspecies *cremoris* (Gobbetti, M, 2000), bioactive peptides acting as enzyme inhibitors (ACE) that influence cardiovascular protection body blood pressure reduction, etc.

High biological value products from sheep's milk is given and fat composition. For this purpose we studied sheep milk in terms of composition of essential fatty acids. Approximate composition of sheep milk fatty acid is the saturated FA (SFA) in milk fat level is quite high (More Than 60%) while monounsaturated FA (MUFA) and polyunsaturated FA (PUFA) has approximately 28% and 6%, respectively (Cabiddu et al.,

2003, 2005). Concentration in sheep milk fatty acids is influenced by many factors: genetics, diet, age, number of lactations, lactation period (A. Cartaica, 2008). Depending on the stage of lactation (Daniel blackbird, 2011), the number of lactations (Mierliță D, 2011) essential fatty acids concentration increases with advancing during these periods. Also this component is used to increase the protein and energy rich food (Mierliță D., 2009). Essential fatty acids ω -3 and ω -6 is very important for the human body: the inflammatory and cardiovascular diseases (ShuWanga, 2009), hepatoprotective (Mohamed Makni, 2010), nerone diseases as: attention disorders in children (Nathan Gadoth, 2008), Parkinson's disease (M. Bousquet, 2009), depression (Sabine Riemer, 2010).

MATERIAL AND METHOD

Sana was from sheep's milk collected during the first period of lactation. It was used from 3000 sheep milk mixture. Lactic culture was used to manufacture Ch Hansen, type CHN 11. In raw milk has been introduced fish oil (purchased from the company "Hofigal") and the mixture was homogenized at 70 ° C and a pressure of 200 bar. in three steps. Next to the classic dupe manufactured sana (Costin GM, 2005, 2007). Depending on the concentration of fish oil in raw milk samples were coded as follows: blank without fish oil: S₀; with the addition of 0.05%: S_{0,05}; with the addition of 0.10%: S_{0,10}; and with the addition of 0.015%: S_{0,15}.

Sensory analyzes were performed by five unauthorized persons.

Physical and chemical analyzes were performed: determination of acidity (according to ISO 6091/2008), the percentage of fat (according S.T.A.S. 6352/2-87).

For fatty acid analysis were performed gas-chromatographic determinations and were detected 19 fatty acids. Working protocol for gas-chromatographic analysis was: milk fat was extracted by using the following protocol: about 1ml of milk samples were mixed with 0,6 ml ammonia 25%, 2ml EtOH, 4ml Ethyl ether and 4 ml hexane and then agitated for 2-3min. After this process the lower layer (the ammonia layer) was discarded. Following this step the mixture was passed through a cellulose filter with Na₂SO₄ and then brought to dryness; transesterification: fatty acids were converted to methyl esters by reaction with boron trifluoride/methanol at 80°C for two hours in a closed Pyrex glass tube. The content was transfered into a separatory funell; the methyl ester extraction: the extraction was made using 10 ml hexane, the hexanic fractions collected were dried using anhydrous sodium sulfate, filtered, concentrated under a nitrogen stream and finally re-eluted in 1 mL hexane. Fatty acids were analyzed by gas chromatography (GC) with flame ionization detection (FID). A 1 μ L sample was injected into the Shimadzu GC-17A series gas-chromatograph, equipped with a 30m polyethylene glycol coated column (Alltech AT-WAX, 0.25mm I.D.,

0.25µm film thickness). Helium was used as the carrier gas at a pressure of 147 kPa. The injector and detector temperatures were set at 260°C. For the oven temperature the following program was used: 70°C for 2 min. then raised to 150°C at 10°C/ min. rate and held at 150°C for 3min., then further raised up to 235°C at a 4°C/min; fatty acids were analyzed by gas chromatography (GC) with flame ionization detection (FID). A 1µL sample was injected into the Shimadzu GC-17A series gas-chromatograph, equipped with a 30m polyethylene glycol coated column (Alltech AT-WAX, 0.25mm I.D., 0.25µm film thickness). Helium was used as the carrier gas at a pressure of 147 kPa. The injector and detector temperatures were set at 260°C. For the oven temperature the following programme was used: 70°C for 2 min. then raised to 150°C at 10°C/ min. rate and held at 150°C for 3min., then further raised up to 235°C at a 4°C/min.

Statistical analysis of data for comparison of samples has been used ANOVA, Tukey and Duncan tests and to compare with blank Dunnet test. To determine the optimal amount of fish oil was used comparing ROC curves (Receiver Operator Characteristic = Operating Characteristic) and Fourier analysis of the correlation type (Sipos, C. et al 2004; Teuşdea A., et al, 2008; Teuşdea A., et al, 2008; Teusdea A, 2009).

RESULTS AND DISSCUSIONS

Sensorial in sana samples without and with fish oil did not taste and smell of fish observed after 7 days at refrigerator temperature maintenance or other foreign taste and odor. In terms of physical and chemical analyzes are not significant differences between samples.

Table 1.

Multiple comparisons for samples S_0 ; $S_{0.05}$; $S_{0.10}$; $S_{0.15}$ sana by Tukey algorithm (HSD) / 95.00% for fatty acids detected chromatographic

Fatty acids	$S_{0.05} \sim S_0$	$S_{0.10} \sim S_0$	$S_{0.15} \sim S_0$	$S_{0.05} \sim S_{0.10}$	$S_{0.05} \sim S_{0.15}$	$S_{0.10} \sim S_{0.15}$
Caproic	0.00223	0.00223	0.00223	1.00000	1.00000	1.00000
Caprilic	0.00223	0.00223	0.00223	0.00223	0.00223	0.00223
Capric	0.00223	0.00223	0.00223	0.00223	0.00223	0.00223
Lauric	0.00223	0.00223	0.00223	0.00223	0.00223	0.00223
Miristic	0.00223	0.00223	0.00223	0.00223	0.00223	0.00223
Miristoleic	1.00000	0.22207	0.00223	0.22207	0.00223	0.00223
Pentadecanoic	0.00223	1.00000	0.00223	0.00223	0.21490	0.00223
Cis-10 pentadecanoic	0.00223	0.00223	0.00223	0.00223	0.00223	0.00223
Palmitic	0.00223	0.00223	0.00223	0.00223	0.00223	0.00223
Palmitoleic	0.00223	0.00223	0.00223	0.00223	0.00223	0.01436
Heptadecanoic	1.00000	0.52570	1.00000	0.53484	1.00000	0.52570
Cis-10 heptadecanoic	0.00223	0.19072	0.19072	0.00434	0.00434	1.00000
Stearic	0.00223	0.14908	0.00223	0.00223	0.00223	0.00223
Oleic	0.00223	0.00223	0.00223	0.00223	0.00223	0.00223
Elaidic	0.00223	0.00223	0.00223	0.00223	0.00223	0.00223
Linoleic ω6	0.00485	0.00485	0.00485	0.00485	0.00485	0.00485
Linolenic ω3	0.00223	0.00223	0.00223	0.00223	0.00223	0.00223
γLinolenic ω6	0.00223	0.00223	0.00223	0.00223	0.00223	0.00223

To validate the statistical variations in samples sana fatty acids as conclusion of all fatty acids detected in the product chromatographic analysis were performed by ANOVA test for multiple comparisons. Table 1 shows these results for centralization and Table 2 Tukey algorithm for iterative algorithm Dunnet.

Table 2.

Multiple comparisons for samples S_0 ; $S_{0.05}$; $S_{0.10}$; $S_{0.15}$ by sana by Dunnet algorithm / 95.00% for the chromatographic fatty acids detected.

Critical range test Dunnet: [-2.89061, +2.89061]

Fatty acids	$S_{0.05} \sim S_0$	$S_{0.10} \sim S_0$	$S_{0.15} \sim S_0$
Caproic	-171.04719	-171.04719	-171.04719
Caprilic	-160.08331	-80.04166	-253.03491
Capric	-252.48038	-76.50921	-172.14571
Lauric	-152.02796	-38.89087	-49.49747
Miristic	28.18855	-20.13468	10.06734
Miristoleic	0.00000	2.13201	-12.79204
Pentadecanoic	10.78328	0.00000	12.93993
Palmitic	237.76806	18.54928	97.80530
Palmitoleic	43.16689	8.22226	12.33340
Heptadecanoic	0.01696	1.41335	0.00000
Cis-10-heptadecanoic	8.98177	2.24544	2.24544
Stearic	-12.12678	2.42536	104.29032
Oleic	249.20798	49.41193	81.63710
Elaidic	23.23790	15.49193	-25.81989
Linoleic ($\omega 6$)	24.82172	56.27038	143.23368
Linolenic ($\omega 3$)	37.94733	79.05694	94.86833
γ -Linolenic ($\omega 6$)	29.39874	53.45225	98.88666

The analysis of surface samples sana chromatograms and fish oil resulting increase their values with increasing concentration of fish oil samples, which shows that the percentage of fatty acids to total fatty acids in the samples increases on addition of fish oil, mainly (Table 3).

Table 3.

Comparison of surface chromatography (ua) of essential fatty acids in sana

$S_{0.05}$	S(u.a.)	$S_{0.10}$	S(u.a.)	$S_{0.15}$	S(u.a.)	Ulei peste	S(u.a.)
Linoleic	41.9709	Linoleic	43.8620	Linoleic	43.6881	Linoleic	64.2705
Linolenic	12.9837	Linolenic	16.1073	Linolenic	15.0795	Linolenic	178.0618
γ -Linolenic		γ -Linolenic		γ -Linolenic		γ -Linolenic	
Linolenic	15.2309		13.5434		12.8500		40.2639

ROC curves use genuine comparison method, we determined the optimal threshold potting essential fatty acids in sana: Figure 1, linoleic acid; Figure 2, linolenic acid; Figure 3, γ -linolenic acid.

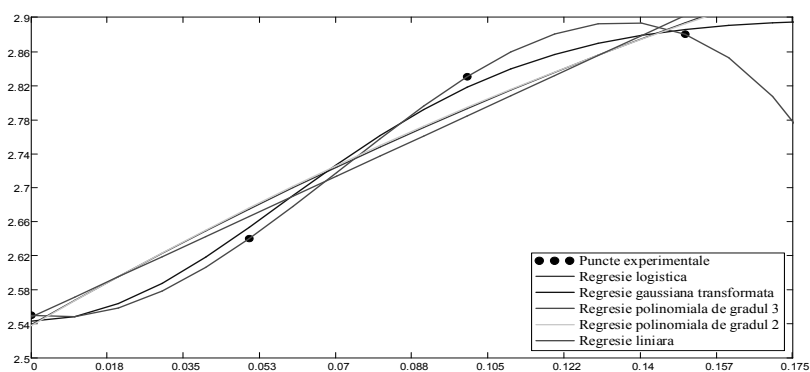


Figure 1. Regression functions studied in the case of linoleic acid of sana

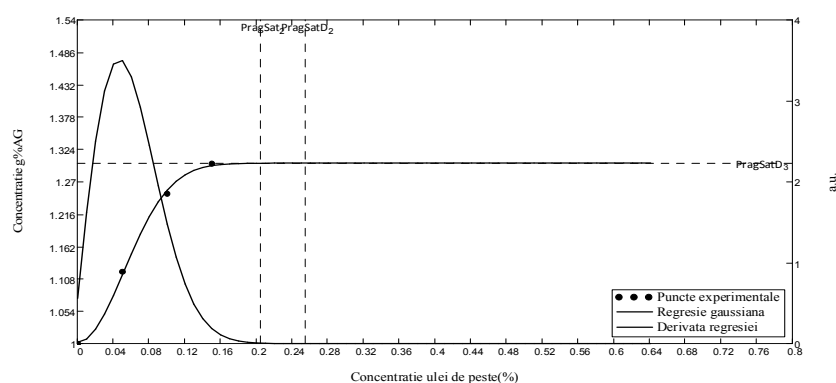


Figure 2. Asymptotic threshold determination linolenic acid potting in sana, with transformed gaussian function

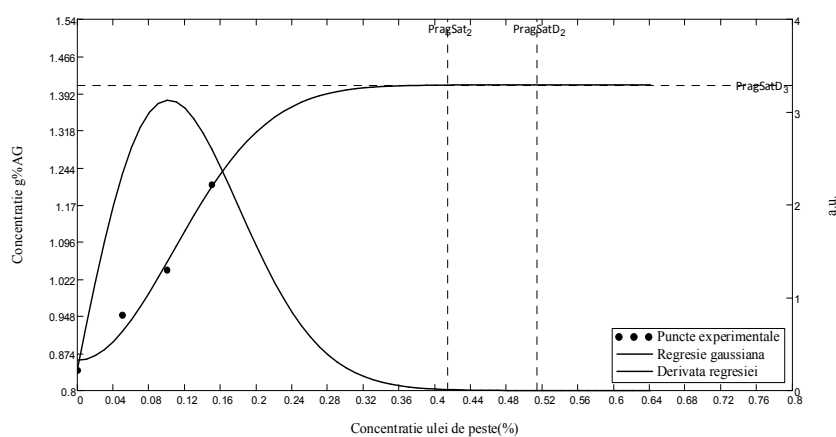


Figure 3. Asymptotic threshold determination potting γ -linolenic acid in sana, with transformed gaussian function

Figure 1 shows that the addition of up to 0.15% fish oil in the milk of manufacture sana potting limit is not reached linoleic acid in fat globule of product. Embed range linolenic acid in fat globule sana is between 0.2 ÷

0.25% (Figure 2), fish oil added to milk and γ -linolenic acid is between 0.4 \div 0.5% . That if linoleic acid is not reached maximum potting may be due to the fact that homogenization was performed first milk and fish oil, followed by pasteurization it is possible that high-fat globule membraba to heat and thus split pass some of the fat globule in plasma.

After centralizing embedding threshold values of fatty acids in fat globule (Table 4), we see that they differ not significantly depending on the function used.

Table 4.

Asymptotic threshold values potting essential fatty acids from sana			
Fatty acids	Embedding threshold (from regression values)	Embedding threshold (the derivative regression values)	Embedding threshold (theoretical)
linoleic	threshold is not reached maximum embedding up to 0.15%		
linolenic	1.301	1.301	0.650
γ -linolenic	1.409	1.409	1.319

By way ANOVA, Tukey test to determine the optimal concentration of fish oil that includes in sana fat globule, according to data.

Table 5.

Fatty acid concentrations to a maximum of embedding the samples sana		
Fatty acids	Conc. of fish oil(%)	Type asymptotic threshold
Linolenic	1.3009	din valorile regresiei
Linolenic	1.3011	din valorile derivatei regresiei
Linolenic	0.6505	teoretic

γ -Linolenic

1.4090 din valorile regresiei

γ -Linolenic

1.4093 din valorile derivatei regresiei

γ -Linolenic

1.3200 teoretic

Table 6.

Number values, sample averages, standard deviation						
Variable	No. value	No. valid values	No. invalid value	sum of weights	MD	SD
Conc. Fish oil (%)	6	6	0	6	1.232	0.289

Table 7.

Multiple comparisons of levels of essential fatty acids sana to embed the maximum produced by Tukey algorithm (HSD) / 95.00%.

Tukey critical Vloarea statistics: 3927.

Categories	Differences	Differences standardized	Critical value	Pr. > Dif.	Significantly
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γ-Linolenic ~						
Linolenic	0.295		1.349	2.776	0.249	Nu
Categories	MD	Groups				
γ-Linolenic						
	1.379	A				
Linolenic	1.084	A				

It is noted that there are significant differences between potting optimal thresholds, so they can consider the average concentration, 1.232% fish oil added in milk, to the theoretical optimal limit potting fish oil in sana.

CONCLUSIONS

The research conducted has shown that sana may be made from sheep's milk with added fish oil. The recommended concentration in terms of technology, is 0.15% lower than that resulting from theoretical view points. Further studies are needed to remove the risk of passing the fat globule fatty acids in plasma by high temperature pasteurization of milk, followed by maintaining the freezing temperatures in order to rehydrate protein and fat cell membranes recovery, possibly split heat, followed by mixing milk with added fish oil. Is necessary and deodorizing mixture of milk with fish oil.

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REFERENCES

1. Cabiddu, A., Decandia, M., Addis, M., Piredda, G., Pirisi, A., Molle, G., 2005, *Managing Mediterranean pastures in order to enhance the level of beneficial fatty acids in sheep milk*, Small Rumin, Res. 59, 169–180.
2. Cabiddu, A., Decandia, M., Molle, G., Pinna, G., Addis, M., Spada, S., Pirisi, A., Piredda, G., 2003, *Effects of different pasture on CLA content in sheep milk and cheese*. In: Mariani, P., Superchi, P., Sabbioni, A., Summer, A. (Eds.), *Proceedings of the ASPA XV Congress*, vol. II. Parma, Italy, pp. 518–520.
3. Cartaa A., Sara Casua, M.G., Usaia, M. Addisa, M. Fiori , A. Fraghia, S. Miaria, Muraa, G., Pireddaa, L. Schiblerc, T. Sechia, J.M. Elsenb, F. Barillet , *Investigating the genetic component of fatty acid content in sheep milk*, Small Ruminant Research, 79 (2008) 22–28.
4. Costin, G. M., Bahrim, G., Borda, D., Curic, M., Florea, T., Hansen, K. F., Popa, C., Rotaru, G., Segal, R., Skriver, A., Stanciu, S. 2005, *Produse lactate fermentate*, Ed. Academica, Galați.

5. Costin, G. M., Cașulschi, T., Pop, D. M., Stanciu, S., Paraschiv, D., 2007, *Produse lactate funcționale*, Ed. Academica, Galați.
6. Gobetti, M., Ferranti, P., Smacchi, E., Goffredi, F., & Addeo, F., 2000, *Production of angiotensin-I-converting-enzyme-inhibitory peptides in fermented milk started by Lactobacillus delbrueckii subsp. Bulgaricus SS1 and Lactococcus lactis subsp. cremoris, FT4*, Applied and Environmental Microbiology, 66(9), 3898–3904.
7. Hanreich Lotte, Edith Zeltner, 2008, *Brânzeturi pentru casă și piață*. pag. 17-19, Editura Zeltner, trad. :Ina MINTICI, Editura M.A.S.T., București.
8. M. Bousquet, C. Gibrat, M. Saint-Pierre, C. Julien, F. Calon, F. Cicchetti, 2009, *Modulation of brain-derived neurotrophic factor as a potential neuroprotective mechanism of action of omega-3 fatty acids in a parkinsonian animal model*, journal homepage, Progress in Neuro-Psychopharmacology & Biological Psychiatry 33, 1401–1408.
9. Mierliță D., C. Maurescu, St. Daraban, F. Lup, 2009, *Effects of energy and protein content in the diet on milk yield and milk fatty acid profile in dairy ewes*, Bulletin USAMV Cluj-Napoca, Animal Science and Biotechnologies, 66(1-2), ISSN 1843-5262, p: 67-73.
10. Mierlita D., Maurescu C, Chereji I., Halma E., Lup F, 2011, *Comparative Study Regarding the Fatty Acids Profile in Sheep Milk Related to the Breed and Parity*, Analele Univversitatii din Oradea, Fascicula: Ecotoxicologie, Zootehnie și Tehnologii de Industrie Alimentară, ISSN 1583-4301.
11. Mierlita Daniel, Elena HILMA, Stelian DARABAN, Florin Lup, 2011, *Influence of Stage of Lactation on Milk Yield and Milk Fatty Acid Profile in Dairy Ewes*, Analele Univversitatii din Oradea, Fascicula: Ecotoxicologie, Zootehnie și Tehnologii de Industrie Alimentară, ISSN 1583-4301.
12. Mohamed Makni, Hamadi Fetoui, El Mouldi Garoui, Nabil K. Gargouri, Hazem Jaber, Jamel Makni, Tahia Boudawara, Najiba Zeghal, 2010, *Hypolipidemic and hepatoprotective seeds mixture diet rich in x-3 and x-6 fatty acids*, Journal homepage: Food and Chemical Toxicology 48, 2239–2246.
13. Muguerza, B., Ramos, M., Sanchez, E., Manso, M. A., Miguel, M., Aleixandre, M. A., et al., 2006, *Antihypertensive activity of milk fermented by Enterococcus faecalis strains isolated from raw milk*, International Dairy Journal, 16, 61–69.
14. Natan Gadoth, 2008, *On fish oil and omega-3 supplementation in children: The role of such supplementation on attention and cognitive dysfunction*, Brain & Development 30, 309–312.
15. Park, Y.W., Haenlein, G.F.W., 2006, *Handbook of Milk of Non-Bovine Mammals*, Blackiston Publishing Professional/Oxford, Ames, Iowa, USA/UK, 449 pp.
16. Sabine Riemer, Michael Maes, Armand Christophe, Winfried Rief, 2010, *Lowered ω -3 PUFAs are related to major depression, but not to somatization syndrome*, journal homepage, Journal of Affective Disorders 123, 173–180.
17. ShuWanga, DayongWub, Nirupa R. Matthana, Stefania Lamon-Favac, Jaime L. Lecker, Alice H. Lichtensteina, 2009, *Reduction in dietary omega-6 polyunsaturated fatty acids: Eicosapentaenoic acid plus docosahexaenoic acid ratio minimizes atherosclerotic lesion formation and inflammatory response in the LDL receptor null mouse*, journal homepage, Atherosclerosis 204, 147–155.
18. Șipoș, C.; Preda, C., 2004, *Statistică economică*, Ed. Mitron, Timișoara.
19. Teușdea, A. C., Modog, T., Mancina A., Drăgan Dan, 2008, *Deformations analisys with fourier correlation*, Annals of DAAAM for 2008 & Proceedins of the 19th International DAAAM Symposium, "Intelligent Manufacturing & Automation: Focus on Next Generation of Intelligent Systems and Solutions", 22-25th October, ISBN 978-3-90150-958-X (ISI Proceedings M/IT).

20. Teuşdea, A.; Modog, T., 2008, *Fourier correlations of dam horizontal movements time series*, Journal of Electrical and Electronics Engineering, ISSN 1844 – 6035, Editura Universităţii din Oradea, Oradea, pg. 267-270.
21. Teuşdea, A.C. & Gabor, G., 2009, *Iris Recognition with Phase-Only Correlation*, Annals of DAAAM for 2009 & Proceedings of the 20th International DAAAM Symposium, ISBN 978-3-901509-68-1, ISSN 1726-9679, pp 690-691, Editor B. Katalinic, Published by DAAAM International, Vienna, Austria.
22. Versiani Matos Ferreira Adaliene, Gleydes Gambogi Parreira, Laura Cristina Jardim Porto, Érica Guihen Mario, Helen Lima Delpuerto, Almir Sousa Martins, Leida Maria Botion, 2008, *Fenofibrate prevents orotic acid—Induced hepatic steatosis in rats*, Life Sciences, Volume 82, Issues 15-16, 876-883.