FORMS OF PRESERVATION OF SOME BOTANICAL ELEMENTS: DIANTHUS SEROTINUS WALDST & KIT., VAR. TRANSILVANICUS NOVÁK

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Abstract

Preservation through in vitro multiplication of the specie Dianthus serotinus Waldst & Kit., var. transilvanicus Novák, was followed, a specie critically threatened (CR), a Dacian-Pannonian endemism, a species from the country's flora, also important for its ornamental value. At the initiation of the in vitro culture we started from the juvenile flower bud, gathered from the field from Alba County (Râpa Roșie, Lancrăm) from young plants at the first flowering. Explants were cultivated on Murashige Skoog (1972) medium with the following variants: $D_o = MS$; $D_I = D_o + 3g/l$ $CV + 1g/l EGP; D_2 = D_o + 0, 1mg/l AIB + 0, 1mg/l BA + 825mg/l NH_4NO_3; D_3 = D_o + 0, 1mg/l AIB + 0, 1mg/l$ $2mg/l BA + 40mg/l AdSO_4$, which after three months of in vitro culture evolved depending on the composition of the medium. The best evolution took place on D_1 (economical medium devoid of phytohormones), on which there were obtained completely conformed plants. On the variants with phytohormones and additional additives there were reported undesired phenomena, vitrification or hyperhidricity of the neoplantlets due to the additional additive of 825 mg/l NH_4NO_3 ; or the differentiation of a weak Radicular System (short roots and few in number), but a large number of neoplantlets, about 85 neoplantlets/explant, due to the presence of BA and of $AdSO_4$ (D_3). Completely conformed neoplantlets (Photo. 1) acclimatized in a percentage of 80%, in compliance with the stages of acclimatization (Laslo, 2013). During the first year they were kept in greenhouses or in cold seedbeds, then in the spring of the second year they were planted in a definitive place (Cluj-Napoca Botanical Garden, Râpa Roșie Reserve, Alba County), a year in which they gained forcefulness. We believe that form of ex situ conservation through in vitro multiplication is in the pipeline and also it is of interest for the conservators and for the species threatened with extinction and which also have an ornamental value. For Dianthus serotinus Waldst & Kit., var. transilvanicus Novák, sozological, being critically endangered specie (CR), Dacian-Pannonian geoelement and endemism, in vitro micropropagation technique makes possible its conservation in a botanical garden, in a reserve or in its area of origin, following periodically the capacity to adapt of the new plantlets at their in vitro transfer.

Keywords: Dianthus serotinus, in situ, ex situ conservation, severely endangered, Dacian element, Dacian-Pannonian endemism, regeneration, in vitro micropropagation, ex vitro acclimatization

INTRODUCTION

In the last two decades there were introduced on the red list over 8300 species of plants with different degrees of endangerment (Sarasan, V et. al., 2006), and numerous natural habitats and plant populations have dropped heavily as number of individuals due to the anthropic activity (Laslo, 2013). According to the IUCN Report from May 2006 the rate of extinction of the species is of 100 up to 1000 times higher than the natural one, therefore appreciating that more than 50% of the species have gone extinct in the last 20 first of all due the climate changes. years, to (http://www.natureserve.org/aboutUs/PressReleases/IUCN_Red_List_release.pdf).. Global Strategy for

Plant Biodiversity (CBD) elaborated at Hague in 2002 within the Convention for Biodiversity, has the task of maintaining the plants' natural patrimony and supports researches in different fields of conservation, establishing preservation protocols of the plant species, also supporting, as main objectives, measures for the *ex situ* conservation (http://www.bgci.org.uk/files/7/0/global_strategy.pdf).

Table 1

No.	Conservation	Form	Categories	
Crt.			_	
А.	"in situ"	Maintenance of the species in their natural ecosystem and habitat	natural 1.Scientific Reserves; 2.National Parks;	
В.	"ex situ"	a. Thematic collections	Active and basic work collections: in Botanical Gardens or in other institutions in the field	
		b. Gene banks	banks In vitro cultures of the seeds and of the tissues; Cryostorage of the germplasm and tissues obtained in vitro at N - liquid temperature	
		c. Multiplication of plants and their conservation through "in vitro" cultures and collections	 In vitro collections with periodical passing and rejuvenation of the material; In vitro cultures with the acclimatization of the <i>ex vitro</i> plants and their relocation in the space of origin 	

Brief presentation of some measures and forms of plant conservation

In 1995 there was established Planta Europa, an organization concerned with the spontaneous flora conservation on the European space, which established the need for long-term conservation of plant diversity, for restoring habitats and ecosystems through programmes and measures presented in Table 1. In situ conservation of plants, according to some researchers, ensures the protection of the species threatened with extinction through the conservation of the biological community and of the ecosystems belonging to them, considered until the middle of the last century as being the only way of conservation (Boşcaiu, 1985). Ex situ conservation is an alternative to the conservation and protection of biodiversity, highlighting the independence between ecology, economy and social life (Seager, 1995), in order to restore de populations of some endangered species through measures taken into another ecological ambiance than the one within the own habitat, through the protection of the endangered elements (Cristea, and Denaever, 2004). Due to the isolation of the tissue cultivated in vitro and to its growth in different conditions towards the ones of the natural life, through in vitro cultures somaclonal variability is induced (Zăpârtan, 1995), that may be due to the method of preservation, to the type of explant, to the medium, genotype, age of the explant donor plant, etc. (Blându and Holobiuc, 2008). After the genomic analysis of the plant material obtained in vitro and after the ex vitro acclimatization there can be initiated repopulation actions of the endangered habitats, after a prior consultation with the specialists from the natural reserves and from the protected areas (Cristea, V. et. al., 2004;). The method of preservation through in vitro collections takes into account the slowing of some growth and multiplication processes (Withers, 1990a), in vitro collections cannot be compared to a gene bank because the material obtained in vitro must be conserved at the highest standards (Laslo, 2011a and b) and even at standards of biological security (Martin and Postman, 1999), and the morphological description of the material must be accompanied, as a general rule, by molecular, biochemical analysis, etc. (Withers, 1990b; Halmagyi and Keul, 2007). The shortcomings of the conservation in gene banks or in clonal field collections made grew interest for *in vitro* conservation, the method becoming certitude, making possible a link between the classical method and the modern one, both of them complementing each other (Engelman, F., 1991; Asmore, 1997), being considered a solution for the conservation of the species that raise issues (Bajaj, 1986; Cachiță et. al., 2007).

Plant species conserved *ex situ* included a wide range of species of economic interest, as the case of *in vitro* obtaining of the planting material at some autochthonous varieties (Agud et. al., 2013; Köteles, 2013), using economical methods of *in vitro* culture (Agud, 2011), or at some ornamental species, as for example, *in vitro* conservation of some collections of roses using repeated subcultures on fresh mediums (Movchan, et. al., 2004), or other species from the spontaneous flora threatened with extinction (*Leontopodiu alpinum, Lilium sp., Fritilaria sp.* etc.) ensuring a higher degree of protection of the plant material thus obtained than of the material obtained *in situ* (Zăpârțan, 1996; 1997;). A large number of *Dianthus* species were conserved through the *in vitro* technique, either from germplasm or from the tissue conserved in gene banks (Holobiuc et. al., 2010), or multiplied *in vitro* and acclimatized for their introduction into the collections (Cristea et. al., 2004; Zăpârțan, 2000) and planted after acclimatization at their place of origin (Zăpârțan, 1994), or following *in vitro* photoautotroph cultures at some *Dianthus* species from the spontaneous flora of the country (Cristea, Victoria 2010).

MATERIAL AND METHOD

Dianthus serotinus Waldst & Kit. specie, var. *transilvanicus* Novák from Caryophyllaceous family, has other synonyms too, depending on the area they populate (Flora RSR). It is a species of scientific interest because it is a Dacian-Pannonian endemism, with a small area and with very poor populations, being a decorative plant, considerations for which it received sozological statute of critically threatened specie (Boşcaiu et. al., 1994). Taxonomically, it is a heliophile plant, perennial, with white or cream flowers, slightly perfumed (Dihoru and Negreanu, 2009). It is a geoelement from the Pannonian basin and the center of Romania, located in several places in Transylvania, in the Olt Defile, etc. (Dihoru and Dihoru, 1994). At Râpa Roșie the specie is protected because here it is located in the reserve, in the other areas *in situ* or *ex situ* conservation measures must be applied; botanical gardens, gene banks, etc. (Fig. 1). Living on a small area and in populations which are poor in specimens, it was resorted to *in vitro* micromultiplication and to the repopulation of the area of origin.



Fig. 1. Mapping areal of *Dianthus serotinus* Waldst & Kit. specie, on the Romanian territory (Dihoru and Negreanu 2009 p. 211)

From the young plant in bud of *Dianthus serotinus* Waldst & Kit., var. *transilvanicus* Novák, isolated from Râpa Roșie and Lancrăm (Alba County) explants were detached consisting of *juvenile buds* which were cultivated *in vitro* on Murashige-Skoog (MS) medium with the following variants: $\mathbf{D}_0 = MS$; $\mathbf{D}_1 = \mathbf{D}_0 + 3g/l \text{ CV} + 1g/l \text{ EGP}$; $\mathbf{D}_2 = \mathbf{D}_0 + 0,1\text{mg/l} \text{ AIB} + 0,1\text{mg/l} \text{ BA} + 825\text{mg/l} \text{ NH}_4\text{NO}_3$; $\mathbf{D}_3 = \mathbf{D}_0 + 0,1\text{mg/l} \text{ AIB} + 2\text{mg/l} \text{ BA} + 40\text{mg/l} \text{ AdSO}_4$.: CV = vegetal coal; EGP = natural extract of corn germ. We regularly watched the evolution of the explants until the organization of true *in vitro* cultures, following over time the capacity of organogenesis and acclimatization of the neoplantlets.

RESULTS AND DISCUTIONS

Following the evolution of the explant formed from juvenile bud detached from *Dianthus serotinus* Waldst & Kit. specie, var. *transilvanicus* Novák, after three months of *in vitro* culture there were observed the following: the regeneration of *in vitro* plantlets took place on the control medium (D_o), in a somewhat smaller number, with a slow but existing evolution; on variant D_1 , the evolution is very good, the presence of the vegetal coal (CV) and also the natural extract of corn germ (EGP) in the administered concentration has a beneficial effect, the medium being also recommended by the economical formula of structure, and also by the lack of phytohormones; on medium D_2 *in vitro* differentiation of neoplantlets takes place in a good number but the phenomenon of hyperhidricity (vitrification) occurs and inhibits plant growth and the formation of a corresponding Radicular System. Vitrification is due to the phytohormones from the medium, associated with a large amount of NH₄NO₃ (825mg/l); - variant D₃ records a good evolution, over 80 neoplantlets/bud of over 1 cm height but with short and frail roots, the addition of AdSO₄ associated with a high dose of BA (2mg/lBA) produced a slight inhibition of the forcefulness of the Radicular System (Table 2).

Climatic factors from the growth chamber were adjusted according to the requirement of the specie, the light intensity with duration of 8 hours light from 24 hours, humidity from the vegetation chamber situated at about 80%, and temperature between 18 -22° C, with automatic adjustment.

The evolution of the juvenile bud detached from *Dianthus serotinus* specie and cultivated *in vitro* (after about 3 months) was followed in fig. 2 and 3 from which we can see that the largest number of neoplantlets was obtained on D_3 (MS+0,1mg/IAIB+2mg/I BA+40mg/I AdSO₄), about 85 plantlets/bud (Fig. 2). The best Radicular System and the best evolution take place on (MS + 3g/I CV + 1g/I EGP) and with the highest capacity of acclimatization (photo 1).

Table 2

Var.	No. pl / length	No. Roots / length	Observations	Bonification
	(cm)	(cm)		
Do	5.5 / 4.0	7.5 / 2.0	Slow evolution	XX
D ₁	15.0 / 4.8	35.0 / 2.0	Very good evolution, very good multiplication	XXXXXX
			and rooting	
D ₂	24.0 / 1.5	6.0 / 1.0	After a good multiplication vitrification occurs	XXXX
			and affects the formation of a good Radicular	
			System, it is necessary to transfer them on a	
			risogen medium	
D3	85.0 / 1.5	12.0 / 0.5	Good evolution, very, very good	XXXXX
			multiplication, less roots, shorter and frailer, it	
			is necessary to transfer them on a simple	
			risogen medium	

The evolution of the juvenile floral bud of *Dianthus serotinus* cultivated on MS medium



Fig. 1. The evolution of the number of plants and of their length at the bud of *Dianthus* serotinus Waldst & Kit., var. transilvanicus Novák (after three months)



Fig. 2 The evolution of the Radicular System at the plantlets of *Dianthus serotinus* Waldst & Kit., var. *transilvanicus* Novák (after three months)

Analyzing the evolution of the bud on the medium variants it is necessary to make a few recommendations in order to find the best and most advantageous formula of medium for obtaining forceful neoplantlets and with a corresponding Radicular System, an essential condition for ensuring *in vitro* acclimatization. Vitrified neoplantlets resulted on variant D_2 , for their good organization they must be transferred on a medium with another dose (1mg/IAIB) without additional NH₄NO₃, or administered alone without cytokinin. Plantlets differentiated on D_3 must be transferred on a medium with auxin in a moderate dose (0,5mg/IAIB) and only with BA or with AdSO₄ (not in mixture).



Photo. 1. The evolution of the bud of *Dianthus serotinus* Waldst & Kit., var. *transilvanicus* Novák (after three months of *in vitro* culture)

CONCLUSIONS

1. *Dianthus serotinus* Waldst & Kit. specie, var. *transilvanicus* Novák from the endemic flora of the country sozologically framed in the critically endangered group (CR), was successfully preserved through in vitro multiplication ensuring the repopulation of some spaces and areas.

2. *Dianthus serotinus* Waldst & Kit. specie, was located at Râpa Roșie and Lancrăm (Alba County), from where explants were harvested for the *in vitro* culture, with the purpose of maintaining the genetic variability within the specie.

3. Totipotency of the tissues detached from the mother plant is the bigger the donor plant of explants is younger, this is why juvenile bud harvested from the areal of the *Dianthus serotinus* Waldst & Kit. species, var. *transilvanicus* Novák., in May was used as explant.

4. The culture medium used was according to Murashige-Skoog with three variants and the control sample. The variant with additional CV and GEP, D_1 ($D_0 + 3g/l CV + 1g/l EGP$), without hormones proved to be the best, this is why we recommend it for an advantageous *in vitro* micropropagation of the specie.

5. On the variants with phytohormones in different doses there were reported undesirable phenomena (vitrification, the lack of roots), in order to end these phenomena, neoplantlets are transferred on a fresh medium with auxin in small concentrations and without cytokinin.

6. Dacian-Pannonian geoelements, endemic for our country, critically endangered, are also preserved *ex situ*, through the *in vitro* multiplication of the specie and through the attempt to repopulate the areas of origin where they can be found in a small areal and with poor populations.

7. The success of *in vitro* multiplication and regeneration depends on the specie, nature and age of the donor plant of explants, each type of tissue having its own capacity of *in vitro* regeneration and multiplication, on the capacity to adapt to the *in vitro* conditions and to resume the metabolic processes of the regenerated neoplantlet.

8. Acclimatization of neoplantlets took place after two years of observations of the capacity to adapt of the material stored in a cold greenhouse or seedbed, therefore the percentage of acclimatization has topped 80%.

REFERENCES

1. Agud, E., 2011, "Economical methods of in vitro tuberization at Solanum Tuberosum L Variety", in: Analele Universității din Oradea, Fascicula: Protecția Mediului,vol.XVI B, Ed. Universității din Oradea, 1-6.

2. Agud E., Zăpârțan, M., Savatti, M., and V. Laslo, V., 2013a, The influence of the moment of sampling of the potato meristem over the in vitro regeneration and diferentiation capacity, in : Bulletin of University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca. Animal Science and Biotechnologies, Vol. 70, Nr. 1, 1843-5262; Electronic, pp. 104-109.

3. Asmore, S.E., 1997, Status Report on the Development and Application of in vitro Techniques for the Conserv. and Use of Plant Genetic Resources, Ed. Engelman, F., IPGRI, Rome, Italy, pp. 1-14

4. Bajaj, Y., 1986, In vitro preservation of genetic resources, IAEA-SM-282/66 Vienna, pp. 43-57

5. Blându, R., I. Holobiuc, 2008, *Conservarea ex situ a speciilor de plante din lista roșie a plantelor superioare în România.*, în: Celui de al XVI-lea Simozion Nați. de Cult. de Țesuturi și Cel. Vegetale., Cluj-Napoca, Ed. Risoprint, pp. 153 – 168

6. Boșcaiu, N., 1985, Criterii pentru constituirea și gestionarea ecologică a resurselor botanice, Ocrti. Nat. Med. înconj., București, 28,2: 126-135

7. Cachiță-Cosma, D., Deliu, C., Ardeleanu, A., 2007, Tratat de biotehnologii vegetale, Vol. II., Ed. Dacia, Cluj – Napoca

8. Cristea, Victoria., M. Miclăuş, M. Puşcaş., C. Deliu, 2004, Conservative micropropagation of some endemic or rare species from the Dianthus L. genus. In: In vitro Cult. and Hortic. Breeding, Fifth IV CHB Symposium Biotehnology, as therorz and Practice in Horticulture, 2004, p. 3-13

9. Cristea, Victoria, 2010, Culturi in vitro fotoautotrofe la speciile de Dianthus endemice și periclitate din România. Rd. Todesco, Cluj-Napoca

10. Cristea Vasile., Denaeyr S., Herreman J.P., Goia, I.,1996, "Ocrotirea naturii și protecția mediului în România", Ed. Univ.Press, Cluj-Napoca

11. Cristea, Vasile, Denaeyer, S., 2004, De la Biodiversitate la OMG-uri, ed. EIKON, Cluj – Napoca 12. Dodds, J.H., 1991, Conservation of plant genetic resources-the need for tissue culture, in: Dodds J.H. (ed.) In vitro metods of conservation of Pl. Genetic Resources, Chapman and Hall, pp.93-111

13. Engelman, F., 1991b, In vitro conservation of horticultural species. Acta Hortic., 298, 327-334

14. Fay, M.E., 1994, In what situations is in vitro culture appropriate to plant conservation? Biodiversity and Conservations, 3, 176-183

15. Halmágyi, A and Butiuc-Keul A., 2007, Conservarea resurselor Genetice vegetale, Ed. Todescă, Cluj – Napoca

16. Holobiuc, I., Mitoi, M., Blându, R., Helepciuc, F., 2010, The establishment of an in vitro gene bank in Dianthus spiculifolius Schur, and D. glacialis ssp. gelidus. II. Medium-term culture characterization in minimal growth conditions. Rom. Biotechn. Lett., 15(2)

17. Köteles N., 2013, Comportamentul in vitro a unor genotipuri de ghizdei (Lotus corniculatus L.) în funcție de balanța hormonală și epoca de prelevare a explantelor, Teză de doctorat, USAMV, Cluj - Napoca

18. Laslo, V., Micropropagarea caisului, Ed. Univ. Oradea, 2007

19. Laslo V., Vicaș S., Agud Eliza, Zăpârțan M., - " Methods of conservation of the plant germplasm. In vitro techniques", în: Analele Universității din Oradea, Fascicula : Protecția Mediului,vol.XVI B, Ed. Universității din Oradea, 2011, ISSN 122-6255,pp.697-708.

20. Laslo, V., Zăpârțan, M. Agud. E., "In vitro conservation of certain endangered and rare species of Romanian spontaneons flora", în: Analele Univ. din Oradea, Fascicula: Protecția Mediului,vol. XVI A, Ed. Univ. din Oradea, 2011, 247-252.

21. Laslo, V., 2013, BIOTEHNOLOGII VEGETALE și aplicațiile lor, Ed. Universității din Oradea

22. Martin, R. R., Postman, J. D., 1999, Phytosanitary aspects of plant germoplasm conservation. In: Benson E.E., (ed.) Plant Conservation Biotehnology, Zaylor and Francis Lrd. London, 63-82

23. Movchan, O., Mitrofanova, O., Klimenko, Z., 2004, Tissue culture application for creation of growing collections of rose in vitro. Growth and development of plants. Theoretical and practical problems. International Scientific Conference. Lithuanian Institute of Horticulture, babtai, 60

24. Murashige T., Skoog, A., 1972, Revised medium for rapid growth and bioassays with tabbacco tissue cultures, Physiol. Plant, 15, pp. 85-

25. Sarasan, V., Cripps, R., Ramsay, M., Atherton, C., McMuchen, M., Prendergast, G., Rowntree, J., 2006, Conservation in vitro of threatened plants – progress in the past decade. In vitro Cellular and Developmental Biol.-Plant, 42, 206-214

26. Seager, 1995, Atlas de la Terre. Le coût écologique de nos modes de vie, la politique des Etats: une vision d'ansamble, Ed. Autrement, paris

27. Zăpârțan, M, 1994, The conservation of some rare and protected plants from Romania using in vitro methods, in: VII-th Intern. Cong. of Plant Tissue and Cell, Culture, Firenze, June 12-17, p. 44

28. Zăpîrțan, M., 1995, Rolul culturilor de țesuturi în conservareaunor specii rare pentru salvarea și extinderea lor în cultură, Contrtibuții Botanice, Univ. Babeș-Bolyai, Cluj-Napoca, pp.217-221

29. Zăpârțan, M, 1996, Conservarea of *Leontopodium alpinum* using *in vitro* culture techiques. Bot. Garden Micropropagation News, Kew, 2, p. 26-19

30. Zăpârțan, M, 1996 Rolul culturilor de țesuturi în conservarea unor specii rare pentru salvarea și extinderea lor în cultură, Contrib. Bot. Cluj – Napoca, p. 217-221

31. Zăpârțan, M, 1997, Fritilaria meleagris L. – specie rară și vulnerabilă conservată prin tehnici de cultură in vitro. În: Actualități și Perspective în Biotehnologiile Vegetale, Ed. Cachița, D., Ardeleanu, A. and Crăciun, C., Ed. Universității "Vasile Goldiș", Arad p. 162-167

32. Zăpârțan, M, 2000, Conservarea florei spontane prin înmulțire in vitro, Ed. ALC MEDIA GROUP, Cluj – Napoca, 135-141

33. Withers, L. A., 1990a, In vitro tehniques for the conservation of crop germoplasm. National Conference on Plant and Animal Biotechnology, Nairobi, Kenya, 1-27

34. Withers, L,A, 1990b, Tissue culture in the conservation of plant genetic resources. International workshop on tissue culture for the conservation of biodiversity and plant genetic resources, Kuala-Lampur, 1-2

** Flora, Republicii Populare Române., T. Săvulescu, (ed.), de la Vol. I din 1952, până la vol. XIII

*** Lista roșie a plantelor superioare din România, Olteanu, M., G. Negreanu., A. Popescu, N. Roman., G. Dihoru., V. Sandală., S. Mihăilescu., 1994, Studii Sinteze Documentații de Ecologie., Academia Română, Institutul de Biol. București

*** Plante rare, periclitate și endemice din flora României – Lista roșie. Ghe. Dihoru și Alexandara Dihoru,1994, Acta Bot. Hort. București,

*** Cartea roșie a plantelor vasculare din România, Dihoru, Ghe., G. Negrean, 2009, Ed. Academiei Române, București

*** Lista roșie a plantelor vasculare dispărute, periclitate, vulnerabile și rare din flora României, Boșcaiu, N., Gh. Coldea, C. Horeanu, 1994, Ocrot. Nat. Med. Înconj., 38 (1)

**** IBPGR (International Broard for Plant genetic resources),1986, Design, planning and operation of in vitro genebanks, IBPGR, Rome

http://www.iucnredlist.org/info/2007RL_Stats_Table%202.pdf

(http://www.bgci.org.uk/files/7/0/global_strategy.pdf).

(http://www.natureserve.org/aboutUs/PressReleases/IUCN Red List release.pdf).