THE OPTIMIZATION OF THE CONDITIONS OF SEPARATING THE SALICYLIC ACID IN HPLC

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Abstract

This paper provide to determined the optimal conditions of separation of the salicylic acid using the cromatographic column (NUCLEOSIL C18), different compositions of the mobile phases (acetonitril, bidiestilled water, methanol, phosphoric acid) and the influence of debit on the separation by HPLC.

Key words: salicylic acid, chromatographic column, mobile phase, HPLC

INTRODUCTION

The salicylic acid is known for its antirheumatic action, which was discovered by Stricker in 1876, and, as a consequence, it has a very wide range of usages.

The sodium salt is used along with other synergic products in healing different rheumatic forms.

Of the worldwide production of salicylic acid, 65% is used for the production of aspirin to obtain salicylates and only 10% for other purposes.

The salicylic acid is obtained through the Kolbe-Schimtt synthesis, which consists of the carboxylation of the sodium peroxide at 125-130 degrees Celsius and at a CO_2 pressure of 7-8 at (Gocan, S., 2002).

It is presented under crystal form with (p.t) of 157-159 degrees Celsius soluble in very hot water, alcohol, ethers, glycerin and partially soluble in benzene and very soluble in chloroform when it is hot.

It is a powerful inhibitor of the pantothenic acid synthesis that is necessary to the life of microorganisms, thus manifesting its antiseptic action. His usage in the conservation of different organic substances is based on this feature. The functional derivatives of the salicylic acid have important anti-inflammatory and analog-antirheumatic properties and they are made through the reaction of either the carboxylic group or the phenolic groups (Gocan, S., 2002).

The properties of the salicylic acid are modified due to the presence of some impurities which appeared in the achieved reaction. These impurities are the 4-hidroxi-izoftalic acid and 4-hidroxi-benzoic (European Pharmacopeea, 1997).

The admitted limits are of 0.1% for the 4-oh-izoftalic acid and of 0,05% for 4-oh-benzoic acid. The specialty literature recommends the usage of high performance liquid chromatography (HPLC) for the separation and UV detection. For this reason, HPLC with UV detection was the method used by us for the elaboration of a through and accurate method for the quantitative analysis of the impurities within the salicylic acid (European Pharmacopeea, 1997).

MATERIALS AND METHODS

The study was conducted in the laboratory of the Department of Analytical Chemistry within the University of Oradea in 2012. For the chromatographic determination we used:

- HPLC purity methanol purchased from the Merck firm;
- HPLC purity acetonitrile purchased from the Merck firm
- Pure phosphoric acid purchased from the Merck firm
- Bi-distilled water prepared in a lab
- Analytic pure salicylic acid
- HPLC Abe & Jasco chromatograph made of:
- Pump module PU-1580
- Gradient unit LG-980-025
- Degasification module DG-980-50
- Manual injector RHEODYNE
- Detector module UV-1575
- Data processing program :BORWING 1.50
- Chromatographic column NUCLEOSIL 100 C18 5µ 10x0,46

The preparation of the testing solution

Due to the presence of 4-OH-benzoic acid and 4-OH-izoftalic acid, as impurities, we have prepared a testing solution with a content of about 0.1% of these substances. For these, we weighed 0.01g of each substance and they were introduced in proportion in a balloon of 10 ml. The substances were dissolved in methyl alcohol and it was brought at the sign with the same solvent. The solution was used for the determination having the aim to optimize the separation conditions.

Carrying out the determination

We worked with a HPLC chromatograph described at point II. The conditions at which the determination were made are: $\lambda = 270$ nm and 237 nm

Debit: 0,5;0,7 1mL/min;

The composition of the mobile phase: acetonitrile:

 $H_2 O + H_3 PO_4 (300+1) = 20:80;30:70$

We injected 20μ L of the solution that was supposed to be analyzed (tested) and the obtained chromatogram was recorded. The data were processed with a BORWING program.

RESULTS AND DISCUTIONS

A. The determination of the optimum wavelength λ :

For the determination of the optimum wavelength determinations were made keeping constant the composition of the mobile phase and debit. (Fig. 1, a and b)



Figure1. The chromatograms obtained for impurities by Salicylic Acid at: a)270nm; b)237 nm

After the analysis made at the 270 nm and at 237 nm, the values obtained were as follows (table 1) for the area of the drops corresponding to the two impurities:

Table 1

Wavelenght	Drops area (µV.sec)						
λ (nm)	Acid 4 – OH- benzoic	Acid 4 – OH- izoftalic					
270 nm	2188377	3508764					
237 nm	1421019	7275189					

The area of the drops corresponding to the two impurities

From the value of the areas and from the chromatograms (fig 1 a,b) we can see that the optimum determination is accomplished at the 270 nm wavelength .

The determination of the optimum (eluent) debit :

We worked with 3 different debits: 0,5;0,7 and 1 mL/min at a wavelength of 270 nm and the acetonitrile mobile phase : $H_2 0 + H_3 PO_4 (300+1) = 20:80$;

The results obtained are presented in table 2:

Table 2

Optimum (eluent) debit										
Debit [mL/min]	Retention time (R _T) [min]		Resolution		Capacity					
	Acid	Acid	Acid	Acid	Acid	Acid				
	4 – OH -	4 -OH-	4 – OH -	4-OH-	4 –OH –	4-OH-				
	benzoic	izoftalic	benzoic	izoftalic	benzoic	izoftalic				
0,5	6,169	10,633	9,13	9,46	615,93	1062,33				
0,7	7,775	10,371	9,55	6,07	776,53	1036,13				
1	3,119	5,328	2,35	8,77	310,87	531,80				

From the results above we can see a very good separation at a debit of 0,5 mL/min but the retaining time is very high, which represents a disadvantage keeping in mind the fact that at a debit of 1mL/min the retaining time is very small, the resolution is very good and the capacity satisfactory.



Figure 2. The chromatogram obtained for debit Figure 3. The chromatogram obtained of the 0,5 mL/min for debit of the 0,7 mL/min



Figure 4. The chromatogram obtained for debit of the 1 mL/min

The fact that the debit of 1mL/min offers the best condition of separation follows from the chromatograms obtained from the 3 debit (fig. 2,3,4)

The determination of the composition of the optimum mobile phase

The separation has been done on a chromatographic column NUCLEOSIL 100 C18 5 μ m 10x10, 46 testing 2 compositions of the mobile phase:

Acetonitril: $H_20 + H_3PO_4 (300+1) = 30:70$ (Fig. 5) Acetonitril: $H_20 + H_3PO_4 (300+1) = 20:80$ (Fig. 6)



mobile phase composition 20: 80

Figure6. *The chromatogram obtained at mobile phase composition 30: 70*

Table 3

	Retention time (R_T)		Resolution	l	Capacity	
Mobile phase	[min]					
composition	Acid	Acid	Acid	Acid	Acid	Acid
	4 - OH-	4 - OH -	4 – OH -	4 – OH -	4 – OH -	4 - OH -
	benzoic	izoftalic	benzoic	izoftalic	benzoic	izoftalic
acetonitril: H ₂ O+ H ₃ PO	3,119	5,328	2,35	8,77	310,87	531,80
(300+1) = 20:80						
acetonitril: H ₂ O+ H ₃ PO	2,571	3,438	2,23	4,41	256,13	342,80
(300+1) = 30:70						

Retention time, resolution and capacity values at different concentration of organic substance in the mobile phase

The data from the table 3 and fig 6. indicate the fact that that a high concentration of organic substance in the mobile phase leads to unsatisfactory results, and the retaining time for a composition of the mobile phase 30:70 is very close, the resolution and the selectivity having small values .

As we can see in figure 5, the optimum composition of the mobile phase is acetonitrile: $H_2O + H_3 PO_4 (300+1) = 20:80$

CONCLUSIONS

From the experimental results obtained we can see that an efficient separation and a fast and accurate quantitative analysis of the salicylic acid impurities is obtained under the following circumstances:

- Detected wavelength of 270 nm;
- Mobile phase composition
- acetonitril: $H_20+H_3PO_4(300+1)=20:80$
- Debit (eluent) of 1mL /min.

REFERENCES

- 1. Gocan, S. (2002) Cromatografie de Înaltă Performanță, Editura Risoprint, Cluj-Napoca.
- 2. European Pharmacopeea (1997) 3rd Edition.
- 3. Oniscu, C.(1988) *Chimia și tehnologia medicamentelor*, Editura Tehnică, București, pp. 332.
- 4. IUPAC (1997), Compendium of Chemical, Terminology, 2nd ed., Blackwell Scientific Publications, Oxford.
- 5. Roman L., Săndulescu R. .(1988) *Chimie analitică*, Vol.3, *Metode separare și analiză instrumentală*, Ed. Didactică și Pedagogică R.A., București.
- 6. Liteanu, C., Gocan S., Hodişan T., Nașcu H.(1974) Cromatografie de lichide Ed. Științifică, București.
- 7. Balaban A. T., Banciu M., Pogany I.(1983) Aplicații ale metodelor fizice în chimia organică Ed. Științifică și Enciclopedică, București.